ISSN: 2319-4820 (Print) 2582-4783 (Online)

Vol 8 Issue 1

#### Review article

# UNDERSTANDING NEUROBIOLOGICAL PATHWAYS OF PAIN REGULATION AND NOVEL THERAPEUTICS FOR PAIN MANAGEMENT

# Bedanta Bhattacharjee\*, Bhargab Deka, Anshul Shakya

Department of Pharmaceutical Sciences, Dibrugarh University, Dibrugarh 786004, Assam, India

#### Abstract

Background: Pain, as unpleasant as it may appear, is a highly vital defense mechanism. It alerts the body to potential or actual injuries or diseases so that preemptive measures can be taken. Noxious stimuli transmit impulses to the spinal cord, which conveys the information to the brain. When nociception fibers detect noxious stimuli on the skin or in interior organs, pain develops. The detection of the signal is taken up by the dorsal horn of the spinal cord and brain stem, which transfers it to various parts of the brain. This route is facilitated by unique substances known as neurotransmitters. Objective: The objective of this review is to understand the role of numerous endogenous neurotransmitters and the regulation of pain processing pathways. Methods: All data were identified, retrieved, and evaluated by searching for peer-reviewed journal articles using Google Scholar and PubMed. Discussion: Neurotransmitters are endogenous chemical messengers, which carry signals from one neuron to another "target" neuron, muscle cell, or glandular cell across a chemical synapse. Some neurotransmitters are excitatory, facilitating message transmission, whereas others are inhibitory, inhibiting transmission. These neurotransmitters play a crucial role in pain regulation and a deeper understanding of the neurobiological pathways could open a doorway for the development of novel therapeutics for pain. **Conclusion:** As a whole, the current review focused on the cellular and molecular processes that support the pain pathway. Furthermore, the pivotal classes of neurotransmitters involved in pain mechanism viz transduction, transmission, and regulation, as well as their locus and potential pharmacological effects, have been thoroughly explored.

**Keywords:** Pain, noxious stimuli, neurotransmitters, cellular mechanism.

# Introduction

Pain is an essential but unpleasant sensation caused by cellular damage or injury. The processes by which noxious stimuli cause animals to feel pain are intricate. Noxious impulses are transduced at the periphery and conveyed to the CNS, where they undergo significant modulation. Finally, the data is sent to the brain, where it is

<sup>\*</sup>Corresponding author's E-mail: bedanta1994@gmail.com

translated into pain [1]. Plasticity can also occur in the pain pathway, and hyperalgesia and allodynia can arise as a result of peripheral and central sensitization. Addressing the adaptability of pain and analgesia in various pain states may help to enhance treatments for the two most common forms of pain, neuropathic and inflammatory pain in which, damage to nerves and tissues causes changes at both the peripheral and central levels [2]. Drugs that act on specific sodium channels in the peripheral nerve may solely target pain-related activity. Peripheral nerve activity may be controlled by agents that act on the peripheral mediators of pain. Cyclo-oxygenase 2 inhibitors, a novel class of non-steroidal antiinflammatory medicines with no gastrointestinal effects, are becoming accessible. The release of peptides and glutamate in the spinal cord activates several receptors, including the glutamate N-methyl-D-aspartate receptor, which works in tandem with the glutamate N-methyl-D-aspartate receptor. Spinal hypersensitivity is caused by other spinal systems. One strategy is to prevent excitability from being generated, although analgesia may be produced by increasing inhibitions. Opioids work by inhibiting central and peripheral C fiber terminals, spinal neurons, and supraspinal processes via presynaptic and postsynaptic inhibitory effects [3]. However, our understanding of pain mechanisms in the brain is relatively limited. Molecular biology and animal models of clinical pain have identified other potential targets, but the idea of a "silver pill" is unlikely. Pain perception involves a huge number of ion channels, receptors, and cell types, and it is hoped that a greater knowledge of them may lead to novel and improved pain treatments.

# The basic mechanism of pain

With the presence of noxious stimuli, the basic mechanism of pain follows three episodes *viz* transduction, transmission, and modulation.

- i. Transduction: Following events occur in the transduction process: At first, the stimulus-induced events are transformed into chemical tissue events. Chemical tissue events, as well as synaptic cleft events, are then translated to electrical events within the neurons. Furthermore, synapses convert electrical impulses in neurons to chemical events [4].
- ii. Transmission: Transmission occurs once transduction is accomplished by communicating the electrical signals inside the neural circuits. However, neurotransmitters present in the synaptic gap are used to convey transmit information from a postsynaptic endpoint of one cell to a presynaptic endpoint of another cell [4].
- iii. Modulation: By up or down-regulation, modulation occurs at all stages of nociceptive pathways *via* the dorsal horn, primary afferent neuron, and higher brain center. All the above-mentioned events lead to the initiation of pain,

which allows us to feel the pain sensation triggered by the stimulus [4]. The basic mechanism of pain is depicted in Fig 1.

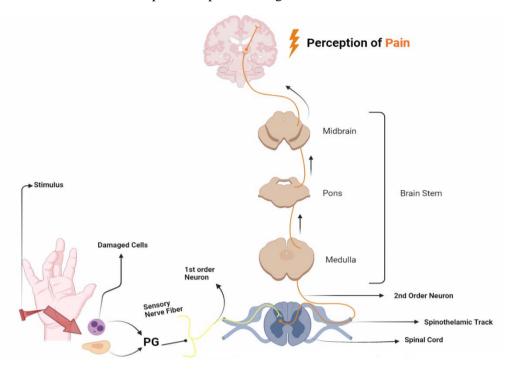


Fig. 1: Depiction of the basic mechanism of pain (Figure created with BioRender.com).

# **Types of Pain**

Based on the mechanisms, symptoms, and syndromes, pain is divided into three categories *viz* inflammatory pain, nociceptive pain, and neuropathic pain.

# **Inflammatory pain**

Inflammation classically describes four key signs. Each of which has a *Latin* derivation. Color or heat, dolor or pain, rubor or redness, and tumor or swelling. Sometimes these four signs combined to cause the fifth sign, which is "function-laesa" or temporary loss of function due to pain or swelling. Inflammation is caused by different types of insults like infections by pathogens, injuries, and exposure to toxins. The ultimate goal of inflammation is to respond to stimuli, protect from harmful agents and restore balance [5]. Oftentimes, that includes eliminating the cause of tissue injury, cleaning out necrotic and dead cells, and starting tissue repair. In this process, inflammation involves activation and sensitization of primary afferent neurons leading to pain sensation. Inflammation can cause three different types of pain responses- allodynia, hyperalgesia, and sympathetic maintained pain.

The localized inflammatory response induces the release of arachidonic acid, which is converted to inflammatory prostaglandins via the cyclooxygenase pathway ultimately leading to pain [5]. Furthermore, inflammation-related pain can be divided into two categories- acute and chronic pain. Inflammatory acute pain is usually severe and lasts only a few minutes, and it is caused by damaging stimuli that are ordinarily transmitted by the A $\delta$ -fibers. Prolonged inflammation, often known as inflammatory chronic pain, lasts longer than expected and is often mediated by C-fibers [6].

# **Nociceptive pain**

Nociceptive pain refers to pain due to an actual or potentially tissue-damaging injury that is transduced and transmitted via nociceptors. The response of our bodies' sensory nerve systems to actual or potentially damaging stimuli is known as nociception or nociperception. Nociceptors are sensory endings that are stimulated by noxious stimuli and are involved in the first phase of the pain perception mechanism [7]. The C-fibers and  $A\delta$  are two different types of primary afferent nociceptors that respond to unpleasant stimuli in our body. Both these two nociceptors are widely located in muscle, skin, bones, joint capsule, and other internal organs as well. They detect potentially damaging mechanical, chemical, and thermal stimuli [7].

# Neuropathic pain

The peripheral nervous system communicates with the rest of the body by connecting nerves in the brain and spinal cord. Signals from the brain could reach the arms, legs, and other internal organs in this way. Neuropathic pain, commonly known as neuropathy or nerve pain, is caused when nerves are injured or destroyed. It causes fatigue, numbness, and moderate to severe pain. Allodynia is frequently accompanied by neuropathy[8]. Allodynia is a type of central pain sensitization caused by repetitive non-painful receptor stimulation. It causes a pain sensation in response to stimuli that are normally non-painful. Diabetes is a common cause of neuropathy, but it can also be caused by injuries, infections, or exposure to specific toxins. Nociceptors exhibit a dynamic expression of ion channels such as voltage-gated sodium channels during neuropathic pain conditions. In addition, voltage-gated sodium channels are important regulators of initiation and propagation of action potentials as well as neuronal excitability [8].

# Role of neurotransmitter in sensitization of pain

The pain perception is mediated by a variety of neurotransmitters including tachykinins, adenosine triphosphate, adenosine, histamine, serotonin, glutamate, leukotriene B<sub>4</sub>, prostaglandin E<sub>2</sub>, prostaglandin I<sub>2</sub>, bradykinin, nerve growth factor, proton, nitric oxide, norepinephrine, Y-aminobutyric acid, calcitonin gene-related

peptide, glycine, and cannabinoids which are summarized in Table 1 along with their expression, receptor mechanism, and pharmacological mechanism.

# **Tachykinins**

Tachykinins are categorized under the neuropeptides family. With its three members family *viz* neurokinin A, neurokinin B, and substance P produced from peripheral terminals of the sensory nerve fibers like skin and muscle can induce neurogenic inflammation through proteolytic cleavage of pre-protachykinins. The neurokinin A, neurokinin B, and substance P bind to their corresponding receptors preferentially based on their affinity. Based on their affinity, neurokinin A binds with neurokinin type 2 receptor, neurokinin B binds to neurokinin type 3 receptor, and substance P binds with neurokinin type 1 receptor respectively. All of the receptors described above are Gq-Protein-coupled receptors, which mediates through phospholipase C/inositol triphosphate and diacylglycerol/protein kinase C signaling pathways upon activation, thereby showing excitatory effects [9].

# **Adenosine Triphosphate**

Adenosine triphosphate is a key intracellular messenger that is liberated locally by the injured tissues and activates its receptors directly. For binding to its receptor, adenosine triphosphate is converted to adenosine through the presence of ectonucleotidases which binds to its receptor i.e ionotropic purino receptor. The ionotropic purino receptor that is of six types shows its action in the region of sensory neurons. The most selectively shown ionotropic purino receptor type 3 receptors are widely expressed in the region of a small C-fibered nociceptor. After, adenosine triphosphate attaches itself to the ionotropic purino receptor type 3 receptors Na<sup>+</sup> ion crosses the channel and because of this membrane depolarizing occurs. Due to this, various Ca<sup>+</sup> sensitive intracellular processes and as a result both pain and hyperalgesia could be perceived. Adenosine triphosphate can promote glutamate release by acting on nociceptors presynaptically. Furthermore, adenosine a by-product of adenosine triphosphate metabolism binds to either adenosine type 1 receptor or adenosine type 2 receptor. Type 1 adenosine receptor is G<sub>i</sub>-protein coupled receptors for inhibitory effects and type 2 adenosine receptor are G<sub>s</sub>-protein coupled receptors which are located both centrally and peripherally sensitize the nociceptor through the cyclic adenosine monophosphate/phosphokinase A pathways [10, 11].

# **Cytokines**

The platelet-activating factor is triggered to release serotonin or 5-HT from circulating platelets during the inflammation-induced mast cell degranulation process. The 5-HT receptors are all G-protein coupled receptors, except for the 5-hydroxytryptamine type 3 receptor, which is a ligand-gated ion channel. The 5-

hydroxytryptamine type 2A receptor and the 5-HT3 receptor are the two principal subtypes of 5-HT receptors found on sensory neuron terminals. Among them, the 5-hydroxytryptamine type 2A receptor is Gq-protein-coupled, which raises the pain sensation via regulating phospholipase C/ inositol triphosphate and diacylglycerol/protein kinase C pathways, while stimulation of 5HT3 receptor causes excitatory effects. Furthermore other cytokines, including TNF- $\alpha$  and IL-1 $\beta$ , also play an important role in causing hyperalgesia by having a significant proinflammatory impact, as well as interacting synergistically with a nerve growth factor [12].

# Glutamate

The amplest excitatory neurotransmitter in the vertebrate system, manifesting itself at peripheral sites of inflammation and contributing to more than 50% of brain synapses. It regulates nociception and the release of neurotransmitters from terminal afferents neurons. In general, sensory neurons exhibit a sufficient number of glutamate receptors, including N-methyl-D-aspartate receptors and alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptors [13].

# Leukotriene B<sub>4</sub>

Leukotriene B<sub>4</sub> is considered an eicosanoid inflammatory mediator formed within the leukocytes by the oxidation of arachidonic acid. After an injury, the arachidonic acid breaks into 5-hydroperoxy eicosatetraenoic acid by the action of enzyme lipooxygenase and thereafter hydrolyzed into leukotriene B<sub>4</sub> by leukotriene A<sub>4</sub> hydrolase. Leukotriene B<sub>4</sub> can elicit hyperalgesia indirectly in the presence of polymorphonuclear leucocytes, most likely *via* the afferent terminal route. By raising the cyclic adenosine monophosphate/phosphokinase A activities, Leukotriene B<sub>4</sub> can induce nociceptors to become more peripherally sensitive [14, 15].

# **Prostaglandins**

Prostaglandins are synthesized from arachidonic acid via the action of the enzyme cyclooxygenase. The two main prostaglandins are PGE<sub>2</sub> and PGI<sub>2</sub>. PGE<sub>2</sub> receptors are classified into 4 types viz prostaglandin E2 receptor type 1-4 (EP1-4), while the receptor of PGI<sub>2</sub> is called prostacyclin receptor (IP). EP<sub>1</sub> is a Gq-protein-coupled receptor that follows phospholipase C/inositol triphosphate diacylglycerol/protein kinase C signaling pathways. EP<sub>2,4</sub> and IP are G<sub>s</sub>-proteincoupled receptors and follow the cyclic adenosine monophosphate/protein kinase A/adenylyl cyclase signaling pathway, whereas EP<sub>3</sub> is G<sub>i</sub>-protein-coupled receptor produces inhibitory effects. Besides this, prostaglandin promotes the action of other chemical mediators like bradykinin and serotonin, along with calcitonin generelated peptide and Substance P to be released. Furthermore, a rise in bradykinin triggers the secretion of prostaglandins and thus produces a "self-sensitizing effect" [16].

# **Bradykinin**

Bradykinin is a protein belonging to the kinin family. Bradykinin is considered an inflammatory mediator is produced locally at the site of inflammation through the fragmentation of large molecules called kininogens. Bradykinin receptor type  $B_1$  or bradykinin receptor type  $B_2$  is Gq- a protein-coupled receptor that binds to bradykinin causing peripheral sensitization in the nociceptive afferent nerve fibers. The activation of bradykinin receptor type  $B_1$  or bradykinin receptor type  $B_2$  leads to activation of phospholipase C to break down the phosphatidylinositol 4,5-bisphosphate into inositol triphosphate and diacylglycerol, further diacylglycerol switch on the protein kinase C, showing a rise in  $Ca^{2+}$  conductance. Moreover, bradykinin can also act synergistically with other substances like nerve growth factor and prostaglandins which further stimulate the formation of pro-inflammatory cytokines [17, 18].

#### **Nerve Growth Factor**

The mediator for chronic pain, locally discharge at the site of abrasion by fibroblast is a neurotrophic factor of neuropeptide known as a nerve growth factor. It can stimulate a rapid onset of thermal and mechanical hyperalgesia by expressing nerve growth factors swiftly after the inflammatory abscess is produced. Tropomyosin receptor kinase which is a nerve growth factor-dependent nerve fiber has a high tendency for nerve growth factor receptors. Nerve growth factor receptor is abundantly expressed in primary afferent neurons, suggesting that it plays a significant role in peripheral sensitization activation. Nerve growth factor, for example, can promote mast cell degranulation and the synthesis of histamine and serotonin, as well as induce the release of additional nerve growth factor, so amplifying inflammatory signals [19].

#### **Proton**

An injured site is generally more acidic than homeostasis thus the protons are more in numbered at the injured site. Around the injured site, a rising number of these protons activates both the vanilloid receptor 1 and acid-sensing ion channels. Moreover, studies have shown that exposing primary afferent nerve fibers to a pH less than 6 can trigger the acid-sensing ion channels [20]. In presence of capsaicin or heat stimuli, this proton can also stimulate vanilloid receptor 1 when pH is less than 5.5 [20]. When both vanilloid receptor 1 and acid-sensing ion channels are activated, the expression of histamine, bradykinin, and prostaglandin E<sub>2</sub> at the injury site further raises the intracellular Ca<sup>2+</sup> influx, thus improves the expression of sensory neuron-specific Na<sup>+</sup> channels and vanilloid receptor 1 [21]. As a result of

the influx of Na<sup>+</sup>, an action potential is a spawn, leading the afferent neurons to become sensitized. The release of calcitonin gene-related peptide and substance P is triggered by an increase in intracellular Ca<sup>2+</sup>, whereas it can also desensitize vanilloid receptor 1 [22].

# Norepinephrine

The primary neurotransmitter of the adrenergic systems, norepinephrine, is produced in nerve terminals from phenylalanine, an essential α-amino acid. An enzyme, tyrosine hydroxylase turns phenylalanine into tyrosine, which is then transformed into 3,4-dihydroxyphenylalanine. 3,4-dihydroxyphenylalanine is subsequently transformed into dopamine, which is the main precursor of norepinephrine that is stored in nerve terminal vesicles. Norepinephrine contains receptors like  $\alpha_1$ - $G_0\alpha$ ,  $\alpha_2$ - $G_1\alpha$ ,  $\beta$ - $G_5\alpha$  protein-coupled receptors. Among them,  $\alpha_1$ - $G_0\alpha$  and  $\beta$ - $G_5\alpha$  are mostly located in postsynaptic neurons, whereas  $\alpha_2$ - $G_1\alpha$  are located in presynaptic neurons. As a result, activation of the  $\alpha_2$ -G<sub>1</sub> $\alpha$  protein-coupled receptors reduces norepinephrine release out of the synapse by blocking Ca<sup>2+</sup> influx. Besides this, the attachment of norepinephrine with  $\alpha_1$ - $G_0\alpha$  and  $\beta$ - $G_s\alpha$  proteincoupled receptors which are present in postsynaptic nerves provoke the phospholipase C/phosphokinase C and cyclic adenosine monophosphate/phosphokinase A signaling pathways, consequently and causes stimulatory effects [23].

# **Y-Aminobutyric Acid**

Υ-Aminobutyric Acid which is considered the most diverse inhibitory neurotransmitter in the mammalian central nervous system contributes nearly 40% of our brain synapses and is present inside the neurons of the cerebellum, spinal cord, and neocortex [24]. Υ-Aminobutyric Acid can interact with ionotropic GABA<sub>A</sub>-receptors or metabotropic GABA<sub>B</sub>-receptors in the nervous system, which are both extensively distributed in the central nervous system, as well as concentrated at presynaptic nerve terminals also. When Υ-Aminobutyric Acid binds to the GABA<sub>A</sub> receptor, the inflow of extracellular Cl<sup>-</sup> into the neurons takes place thereby reducing membrane potential thus producing an inhibitory effect. Moreover, Υ-Aminobutyric acid-binding to GABA<sub>B</sub> receptors, inhibits cyclic adenosine monophosphate production, since the GABA<sub>B</sub> receptor is a G<sub>i</sub>-protein-coupled receptor [25].

# **Calcitonin Gene-Related Peptide**

Calcitonin Gene-Related Peptide is synthesized in both the peripheral and nervous systems, however, it is found largely in the primary afferent nerves. Calcitonin Gene-Related Peptide mimics the excitatory effects of substance P, produces the release of Ca<sup>2+</sup> ions, and is associated with the transmission of noxious stimulation

[26]. The receptors present in Calcitonin Gene-Related Peptide are Gs-protein-coupled which indicates Calcitonin Gene-Related Peptide-mediated pain transmission [27].

# **Cannabinoids**

The cannabinoid is a type of neurotransmitters that attaches themselves to their receptors and regulates the release of neurotransmitters in the brain. Cannabinoids attach to G<sub>i</sub>-protein coupled cannabinoid type 1 receptors which are abundantly expressed in the presynaptic and postsynaptic region of the brain and spinal cord, along with G<sub>i</sub>-protein-coupled cannabinoid type 2 receptors, which are mostly found in the immune system [28]. The expression of cannabinoid type 1 receptor and cannabinoid type 2 receptors suppressed the formation of intracellular cyclic adenosine monophosphate, thus produces a tremendous reduction of the stimulatory effect within the neurons. Furthermore, cannabinoid type 2 receptor activation can inhibit degranulation of mast cells and the production of pro-inflammatory mediators, making a decrease in pain sensation drastically and effectively [29].

# **Opioid Peptides**

Binding of opioid receptors viz  $\delta$ -opioid receptors,  $\kappa$ -opioid receptors, and  $\mu$ -opioid receptors with the peptides are collectively called opioid peptides. All opioid receptors are classified under  $G_i$ -protein-coupled receptor, which implies that when they are activated, they suppress cyclic adenosine monophosphate/adenylyl cyclase activity. The opioid receptors are predominantly found in both the regions of postsynaptic neurons dendrites and primary afferent neurons, and there are two opioid peptides viz dynorphin and enkephalin produced endogenously into the interneurons of the dorsal horn. Such peptides block the transmission of excitatory neurotransmitters from the afferent nerve terminals, thereby attenuating the excitability of neurons and overall controls the pain sensation [30].

Table 1: Neurotransmitters along with their expression and pharmacological mechanism

Neurotrans mitters	Expression of neurotransmitte rs	Receptors: Mechanis m	Inhibitory/ Excitatory effects	Pharmacological Refere mechanism nces	e		
Inflammatory mediators							
Tachykinins- Neurokinin A, neurokinin	Peripheral nervous system ( particularly in C- fibers) and	Neurokinin type 1 receptor, Neurokinin	Excitatory	Mediates neurogenic [9, 31] inflammation; activation of nitric oxide synthase and arachidonic acid			

	G . 1			1 6 1 1
B, and	Central nervous	type 2		pathways for the release of nitric oxide and
substance P	system	receptor,		
(Neuropeptid	(predominantly in the dorsal horn of	and Neurokinin		prostaglandin E2
es)				respectively; improve Cyclic adenosine
	the spinal cord)	type 3		•
		receptor:		monophosphate/protein kinase A activities
		Phospholip ase		Killase A activities
		C/Inositol		
		triphosphat		
		e,		
		Diacylglyc		
		erol/Protein		
		kinase C		
Adenosine	Peripheral	Purino	Excitatory	Improve glutamate [10,
triphosphate	nervous system	receptor-	(Purino	release; sensitize the 32]
and	and Central	$Na^+/K^+$ ; $A_1$	receptor	nociceptors
adenosine	nervous system	and $A_2$ -	and $A_2$ )/	
(Purine)		Adenylyl	Inhibitory	
		cyclase/Cy	$(A_1)$	
		clic		
		adenosine		
		monophosp		
		hate/Protei		
		n kinase A		
Histamine	Peripheral	Histamine <sub>1</sub> -	Excitatory	Exert synergistic [33,
(Monoamine	nervous system	Phospholip	Excitatory	interaction with nerve 34]
)	and Central	ase		growth factor
,	nervous system	C/Inositol		gro war raccor
		triphosphat		
		e,		
		Diacylglyc		
		erol/Protein		
		kinase C		
Serotonin	Peripheral	5-	Excitatory	Exert synergistic [35,
	nervous system	hydroxytry		interaction with nerve 36]

	and Central nervous system	ptamine type 2A receptor- Phospholip ase C/Inositol triphosphat e, Diacylglyc erol/Protein kinase C; 5- hydroxytry ptamine type 3 receptor- Na <sup>+</sup> /K <sup>+</sup>		growth factor		
Glutamate (amino acid)	Peripheral nervous system (particularly in C- fibres) and Central nervous system	N-methyl-D-aspartate receptors and Amino-3-hydroxy-5-methyl-4-isoxazolepr opionic acid receptor-Mg <sup>2+</sup> /Ca <sup>2+</sup> /Na <sup>+</sup> /K <sup>+</sup>	Excitatory			[37]
Leukotriene B <sub>4</sub>	Peripheral nervous system	Leukotrien e B <sub>4</sub> type 1 receptor and Leukotrien e B <sub>4</sub> type 1 receptor 2-	Excitatory/ Inhibitory	Promote production; nociceptors; neutrophils to in	cytokine sensitize recruit jury site	[38]

# Bhattacharjee et al.

Adenylyl cyclase/Cy clic adenosine monophosp hate/Protei n kinase A or Phospholip ase C

Prostagland

Excitatory

(Prostacycli in E2 receptor n receptor, type 1-Prostagland Phospholip in E2 receptor ase C/Inositol type 1, Prostagland triphosphat e, in E2 Diacylglyc receptor erol/Protein type 2, and kinase Prostagland C: Prostagland in E2 in E2 receptor receptor type 4e); type 2, Inhibitory Prostagland (Prostaglan din E2 E2 in receptor receptor type 3) type 3, Prostagland in E2 receptor type 4, and Prostacycli n receptor-Adenylyl cyclase/Cy

Augment the release of [39, bradykinin, calcitonin 40] gene-related peptide, histamine, interleukin-2, serotonin, and substance P

clic
adenosine
monophosp
hate/Protei
n kinase A

Bradykinin (Neuropeptid e)	Peripheral nervous system and Central nervous system (hypothalamus and pituitary)	Bradykinin receptor type B1 and Bradykinin receptor type B2-Phospholip ase C/Inositol triphosphat e, Diacylglyc erol/Protein kinase C	Excitatory (Bradykini n receptor type B1 and Bradykinin receptor type B2)	Exert synergistic interaction with nerve growth factor and prostaglandin; augment the release of prostaglandin, nerve growth factor, and proinflammatory cytokines <i>i.e</i> Interleukin2	[41]
Nerve growth factor (Neuropeptid e)	Peripheral nervous system and Central nervous system	Tropomyos in receptor kinase A- Phosphoino sitide 3- kinase	Excitatory	Augment the release of serotonin, histamine; cause the mast cells degranulation	[42]
Proton	Peripheral nervous system and Central nervous system	Acid- sensing ion channels and Vanilloid receptor 1- Na <sup>+</sup> /K <sup>+</sup>	Excitatory	Improve the release of bradykinin, substance P, calcitonin gene-related peptide, histamine, and prostaglandin E2	[43, 44]
Nitric oxide (Gasotransmi	Peripheral nervous system	Soluble guanylyl	Excitatory/ Inhibitory	Recruited to the site of inflamed tissue	[45]

# Bhattacharjee et al.

tter)	and nervous s	Central system	cyclase/ Cyclic guanosine monophosp hate		
Norepinephri ne (Monoamine )	Periphera nervous and nervous s	system Central	α1- Phospholip ase C/inositol triphosphat e, Diacylglyc erol/Protein kinase C α2- Adenylyl cyclase/Cy clic adenosine monophosp hate/Protei n kinase A	Excitatory ( $\alpha 1$ and $\beta$ )/ Inhibitory ( $\alpha 2$ )	[23]

Non-inflammatory mediators					
Υ-	Peripheral	Υ-	Inhibitory		[46]
aminobutyric	nervous system	•	(Υ-		
acid (Amino acid)	and Central nervous system	ic acid type A receptor-	aminobutyr		
	•	Cl <sup>-</sup> /K <sup>+</sup>	ic acid type		
		(Inhibitory	A receptor		
		postsynapti c	and $\Upsilon$ -		
		potentials);	aminobutyr		
		Υ-	ic acid type		
		aminobutyr ic acid type	B receptor)		
		B receptor-			
		Adenylyl			
		cyclase/Cy			

		clic adenosine monophosp hate/Protei n kinase A			
Calcitonin gene-related peptide (Amino acid)	Peripheral nervous system and Central nervous system (predominantly in the dorsal horn of spinal cord)	Calcitonin receptor- like receptor- Adenylyl cyclase/Cy clic adenosine monophosp hate/Protei n kinase A	Excitatory	Synergistic with the excitatory effect of substance P	[47, 48]
Glycine (Amino acid)	Central nervous system	Glycine receptor- Cl <sup>*</sup> (Inhibitory postsynapti c potentials)	Inhibitory		[49]
Cannabinoid s (Lipid)	Peripheral nervous system and Central nervous system (brain)	Cannabinoi d type 1 receptor and cannabinoi d type 2 receptors- Adenylyl cyclase/Cy clic adenosine monophosp hate/Protei	Inhibitory (Cannabino id type 1 receptors and cannabinoi d type 2 receptors)	•	[50, 51]

# n kinase A

n kinase A	Opioid peptides (Neuropeptid e)	Peripheral nervous system and Central nervous system (hippocampus, hypothalamus, spinal cord, and striatum)	δ-opioid receptors, κ-opioid receptors, and μ-opioid receptors-Adenylyl cyclase/Cy clic adenosine monophosp hate/Protei n kinase A	Inhibitory		[52]
------------	---------------------------------	---	--	------------	--	------

# Novel therapeutic platform for the management of pain relief

By establishing new molecular targets and novel ways to treat pain, we may indeed be able to develop novel medications to effectively treat acute and chronic inflammatory pain without the downsides of present opioid medications. Table 2 and Table 3 illustrated the pain related target along with their mechanism of action, and venom derived peptide used in pain therapy.

Table 2: Molecular target, activators/inhibitors, mechanism of action, and concluding remarks

Molecular target		Activators/Inhibitors	Mechanism of	Concluding	References
			action	remarks	
Opioid	receptor-	Activators-	Supraspinal	Completed phase I	[53]
like	receptor	Cebranopadol,	stimulation has a	trial and	
1/Nocice	eptin	MCOPPB, and	pronociceptive	continuing phase II	
receptor		NNC63-0532;	effect, while	and III clinical	
		Inhibitors- JTC-801,	spinal and	trials, but no	
		LY-2940094, and SB	peripheral	findings have been	
		61211.	stimulation has	published.	
			an anti-		
			nociceptive		
			effect.		

Kappa opioid receptor	Activators- CR 665, CR 845; Inhibitors- Buprenorphine.	Anti-nociceptive activity is primarily peripheral; opioid-induced hyperalgesia is linked to spinal activation.		[54, 55]
Opioid heteromers	Activators- N-naphthoyl-β-naltrexamine; Inhibitors- CYM51010.	Cannabinoid receptor type 1 and mu-opioid receptor heteromers are colocalized in the rostral ventromedial medulla and dorsal root ganglion; mu- opioid receptor heteromer found in dorsal root ganglion and rostral ventromedial medulla.	Initial phases of development.	[56]
Serotonin, norepinephrine, and dopamine	Activators- Antidepressant; Inhibitors- Duloxetine, milnacipran, and venlafaxine.	Inhibit descending pathway.	Serotonin and norepinephrine reuptake inhibitors and tricyclic antidepressants have strong evidence of efficacy in	[57, 58]

peripheral neuropathic pain disorders: serotonin and norepinephrine reuptake inhibitors have evidence of efficacy in nociceptive pain; Serotonin and norepinephrine reuptake inhibitors have minimal evidence of efficacy.

Alpha 2 receptor

Activators- Clonidine, tizanidine, and dexmedetomidine.

Pre- and postsynaptic pain processing are suppressed by anti-nociceptive activity in the spinal region.

Administration *via* [59] intrathecal route may be helpful in mixed or neuropathic pain conditions; clinical evidence suggests modest long-term pain reduction.

Histamine receptor

Activators- Immepip; Inhibitors- GSK 189254, thioperamide.

Histamine release is inhibited by H3 receptor activation; Substance P and Calcitonin generelated peptide release is reduced by peripheral H3 receptor activation;

Preclinical data for analgesia in H3 receptor-activated nociceptive pain models; preclinical data in neuropathic pain models with brain-penetrating H3 receptor antagonists.

nociceptive transmission is reduced by intracerebral histamine signaling.

Cannabinoid receptors

Activators- AM 1241.

Cannabinoid receptors type 1 on the presynaptic membrane impaired neurotransmitter release while Cannabinoid receptors type 2 receptors are activated in neuropathic pain models.

Cannabinoids have [61] strong evidence for treating neuropathic pain and limited evidence for treating nociceptive pain, but they have a high rate of side effects; preclinical evidence of cannabinoid receptors type 1 receptor agonists in neuropathic and nociceptive pain models, but negative findings in clinical studies; preclinical evidence of cannabinoid receptors type 2 receptor agonists in neuropathic and nociceptive pain models, but no

clinical data.

Voltage-gated Activators- Ca<sup>2+</sup>; In the central and N-type channel [62]

# Bhattacharjee et al.

calcium channels	Inhibitors- Mibefradil, gabapentin, ethosuximide, and Z944	peripheral neurological systems, voltage- gated calcium channels regulate neurotransmitter release, ion conductance, and neuronal excitability.	blocking agent has preclinical efficacy in neuropathic pain, but clinical data are lacking; T-type channel blocking agent has preclinical efficacy in neuropathic and nociceptive pain models; clinical trials are still going on, but no findings have been released yet.	
Voltage-gated sodium channels	Activators- Na <sup>+</sup> , brevetoxin B, and ciguatoxin; Inhibitors- TV-45070, CNV1014802, and PF-05089771.	In neurons, it is in charge of the initiation and propagation of action potentials.	Voltage-gated sodium channels 1.7 inhibition has been shown in preclinical studies to alleviate neuropathic and nociceptive pain; Preclinical evidence for voltage-gated sodium channels 1.8 inhibition in neuropathic and nociceptive pain models.	[63, 64]
N-methyl-D- aspartate receptor	Activators- N-methyl-D-aspartate, glutamate, glycine, and Ca <sup>2+</sup> ; Inhibitors- Traxoprodil, ifenprodil,	N-methyl-D- aspartate receptor implicated in central	Clinical data for lower affinity antagonists is conflicting; there is modest	[65, 66]

dextromethorphan, and sensitization

preclinical

Transient receptor potential V1/Vanilloid receptor 1	Activators- Resiniferatoxin, capsaicin; Inhibitors- A784168, AMG517,A795614, and AMG 9810	(both short and long term); N-methyl-D-aspartate receptor is ionotropic. glutamate receptors that are activated by severe or persistent nociceptive stimuli. Transient receptor potential V1 channels are desensitized by agonists; transient receptor potential V1 are ion channels that are sensitive to membrane depolarization, temperature (>42°C), pH (6), and a wide range	analgesia with N-methyl-D-aspartate receptor subunit 2B antagonists; glycine B antagonists have been proven in preclinical investigations to be effective for neuropathic pain.  There is substantial evidence to support the use of topical agonists in the treatment of neuropathic pain; in preclinical research, antagonists give evidence for nociceptive and neuropathic pain; although clinical studies have been	[67, 68]
Mass oncogene- related gene	Activators- BAM8-22, Tyr6-γ2-MSH-6-12.	of ligands.  Mass oncogenerelated gene	completed, no outcomes have been published.  Evidence from nociceptive and	[69]
receptor		receptor activation suppresses postsynaptic	neuropathic pain models in the preclinical stage.	

currents	in	the	
substantia	a		
gelatinos	a	via	
blocking	N-	type	
calcium channels			
in the do	rsal	root	
ganglion.			

Nerve growth Inhibitors- Tanezumab, factor inhibitors fasinumab, ABT110, and REGN-475.

Nerve growth factor binds to tropomyosin receptor kinase A receptors on nociceptive nerve terminals, causing enhanced hypersensitivity and pain signaling.

In the neuropathic and nociceptive pain model, there is strong clinical evidence for humanized antinerve growth factor antibodies.

Tissue necrosis Inhibitors- Infiximab, factor- $\alpha$  inhibitors etanercept.

During pain sensation, tissue necrosis factor-α triggers the growth factors and proinflammatory cytokines mediators.

In neuropathic and [71] nociceptive pain model, strong evidence for tissue necrosis factor-α; supporting strong evidence against rheumatologic condition during the clinical stage.

Glial cell Inhibitors- SB203580, inhibitors CEP-1347, D-JNKI-1, and propentifylline.

Peripheral nerve damage triggers glial cells, which produce inflammatory mediators that make In neuropathic [72] pain model, supporting strong preclinical evidence for glial cell inhibitor.

nociceptive neurons in the area more sensitive.

Nitric oxide and phosphodiesterase inhibitors	Inhibitors- NCX701.	The exact mechanism of pain sensation is unknown, though it may be dependent on the cyclic guanosine monophosphate signaling pathway being activated; analgesia from nonsteroidal anti-inflammatory drugs and opioids is enhanced.	findings when phosphodiesterase type 5 inhibitors are used with morphine; in preclinical models, fewer stomach lesions were	[73]
Bisphosphonates		The analgesic mechanism is unknown; it may work by inhibiting osteoclasts and having anti-inflammatory effects	pain syndrome with low-quality evidence; strong	[74]

Table 3: Toxins used as pain therapy in development.

Toxin	Species	Molecular	Clinical	Manufacturer	Reference
		target	stage		S
Conantokin -G (CGX- 1007)	Conus geographu s	N-methyl-D- aspartate receptors, N- methyl-D- aspartate receptor subunit 2B	Phase II	Cognetix Inc.	[75]
Contulakin- G (CGX- 1160)	Conus geographu s	Neurotensin receptor	Phase II	Cognetix Inc.	[75]
ω- Conotoxin CVID (AM336)	Conus catus	N-type Ca <sup>2+</sup> channels	Phase II	Zenyth Therapeutics	[76]
ω- conotoxin MVIIA (Prialt <sup>TM</sup> )	Conus magus	N-type Ca <sup>2+</sup> channels	Approve d by FDA	Elan	[77]
α- Conotoxin Vc1.1 (ACV1)	Conus victoriae	Neuronal nicotinic acetylcholin e receptors	Phase I	Metabolic Pharmaceutica Is	[78]
χ- conotoxins Mr1A/Mr1 B (Xen2174)	Conus marmoreu s	Neruronal noradrenalin e transporter	Phase I/IIa	Xenome	[79]
ρ- conotoxin T1A	Conus tulipa	α1- adrenoceptor s	At preclinica l stage	Xenome	[75]

#### Conclusion

A major problem for the future of pain treatment is the current gap between fundamental scientific research and the development of novel pain therapeutics. This conundrum is particularly challenging to solve when trying to find improved treatments for comorbid chronic pain conditions because: a) Animal 'pain' models may not properly reflect complex clinical pain; b) Subjective pain experience is not assessed in animal behavioral testing; c) Preclinical data are inconclusive when it comes to the direction of novel analgesic research; d) Clinical trials frequently use overly-sanitized research participants, failing to capture the multifaceted effects of comorbid chronic pain. To recapitulate, pain research and management require a thorough grasp of the complicated mechanisms of pain. As a whole, the current review focused on the cellular and molecular processes that support the pain pathway. Furthermore, the pivotal classes of neurotransmitters involved in pain mechanism viz transduction, transmission, and regulation, as well as their locus and potential pharmacological effects, have been thoroughly explored. This could help worldwide researchers gain a better grasp of the pain topic and provide a useful and effective roadmap for future analgesic drug discovery.

# **Future perspective**

Pain is seen as quite unpleasant, and it can be debilitating at times. Pain affects one's quality of life and limits productivity at work. As a result, pain therapy or management is critical in this context. Despite significant progress has been made in understanding neurobiological aspects of pain, more sophisticated research using modern tools and approaches is still required, which may pave the way for a deeper understanding of neurobiological pathways and the development of innovative therapeutics for pain management.

**Conflict of Interest:** The authors declare no conflict of interest.

# Funding: Nil References

- 1. Frangos, E., E.A. Richards, and M.C. Bushnell, *Neurobiology of Pain*.
- 2. Bell, A., *The neurobiology of acute pain*. The Veterinary Journal, 2018. **237**: p. 55-62.
- 3. Price, T.J. and M.S. Gold, From mechanism to cure: renewing the goal to eliminate the disease of pain. Pain medicine, 2018. **19**(8): p. 1525-1549.
- 4. McCarthy, C.J., R. Vazquez, and T.G. Walker, *Specialty Imaging: Acute and Chronic Pain Intervention E-Book.* 2020: Elsevier Health Sciences.
- 5. Yoo, S.K., et al., *Lyn is a redox sensor that mediates leukocyte wound attraction in vivo*. Nature, 2011. **480**(7375): p. 109-112.

- 6. Basbaum, A.I., et al., *Cellular and molecular mechanisms of pain.* Cell, 2009. **139**(2): p. 267-284.
- 7. Kaliyaperumal, S., et al., *Animal models of peripheral pain: Biology review and application for drug discovery.* Toxicologic pathology, 2020. **48**(1): p. 202-219.
- 8. Li, Z.-H., et al., Cyclic nucleotide signaling in sensory neuron hyperexcitability and chronic pain after nerve injury. Neurobiology of Pain, 2019. **6**: p. 100028.
- 9. Donkin, J.J., et al., *Substance P in traumatic brain injury*. Progress in brain research, 2007. **161**: p. 97-109.
- 10. Jarvis, M.F., Geoffery Burnstock's influence on the evolution of P2X3 receptor pharmacology. Purinergic Signalling, 2020: p. 1-7.
- 11. Illes, P., et al., *Update of P2X receptor properties and their pharmacology: IUPHAR Review 30.* British Journal of Pharmacology, 2021. **178**(3): p. 489-514.
- 12. Okamoto, K., et al., 5-HT2A receptor subtype in the peripheral branch of sensory fibers is involved in the potentiation of inflammatory pain in rats. Pain, 2002. **99**(1-2): p. 133-143.
- 13. Meldrum, B.S., *Glutamate as a neurotransmitter in the brain: review of physiology and pathology.* The Journal of nutrition, 2000. **130**(4): p. 1007S-1015S.
- 14. Yam, M.F., et al., General pathways of pain sensation and the major neurotransmitters involved in pain regulation. International journal of molecular sciences, 2018. **19**(8): p. 2164.
- 15. Takeda, T., et al., Dioxin-induced increase in leukotriene B4 biosynthesis through the aryl hydrocarbon receptor and its relevance to hepatotoxicity owing to neutrophil infiltration. Journal of Biological Chemistry, 2017. 292(25): p. 10586-10599.
- 16. Smith, D.L., Cyclooxygenase and lipoxygenase products: A compendium, in CRC Handbook of Eicosanoids: Prostaglandins and related lipids. 2017, CRC Press. p. 47-83.
- 17. Loh, Y.C., et al., Overview of antagonists used for determining the mechanisms of action employed by potential vasodilators with their suggested signaling pathways. Molecules, 2016. **21**(4): p. 495.
- 18. Loh, Y.C., et al., Overview of the microenvironment of vasculature in vascular tone regulation. International journal of molecular sciences, 2018. **19**(1): p. 120.

- 19. Price, M.P., et al., *The DRASIC cation channel contributes to the detection of cutaneous touch and acid stimuli in mice*. Neuron, 2001. **32**(6): p. 1071-1083.
- 20. Gould III, H.J., Complete Freund's adjuvant-induced hyperalgesia: a human perception. Pain, 2000. **85**(1-2): p. 301-303.
- 21. Rajan, S., et al., Ca2+ Signaling by TRPV4 Channels in Respiratory Function and Disease. Cells, 2021. **10**(4): p. 822.
- 22. Chahl, L.A., *The Effect of the Trpv1 Agonist, Capsaicin, on the Developing Rat Brain: A Mini Review.* Journal of Neurology & Neuromedicine, 2020. **5**(1).
- 23. Alluri, S.R., et al., *PET Radiotracers for CNS-Adrenergic Receptors: Developments and Perspectives.* Molecules, 2020. **25**(17): p. 4017.
- Watanabe, M., et al., *GABA and GABA receptors in the central nervous system and other organs.* International review of cytology, 2002. **213**: p. 1-47.
- 25. Papasergi-Scott, M.M., et al., *Structures of metabotropic GABA B receptor*. Nature, 2020. **584**(7820): p. 310-314.
- 26. Chen, L.-J., et al., Expression of calcitonin gene-related peptide in anterior and posterior horns of the spinal cord after brachial plexus injury. Journal of Clinical Neuroscience, 2010. **17**(1): p. 87-91.
- 27. Poyner, D.R., et al., *International Union of Pharmacology. XXXII. The mammalian calcitonin gene-related peptides, adrenomedullin, amylin, and calcitonin receptors.* Pharmacological reviews, 2002. **54**(2): p. 233-246.
- 28. Pacher, P., S. Bátkai, and G. Kunos, *The endocannabinoid system as an emerging target of pharmacotherapy*. Pharmacological reviews, 2006. **58**(3): p. 389-462.
- 29. Demuth, D.G. and A. Molleman, *Cannabinoid signalling*. Life sciences, 2006. **78**(6): p. 549-563.
- 30. Marbach, F., et al., *Variants in PRKAR1B cause a neurodevelopmental disorder with autism spectrum disorder, apraxia, and insensitivity to pain.* Genetics in Medicine, 2021: p. 1-9.
- 31. Szymaszkiewicz, A., et al., *The place of tachykinin NK2 receptor antagonists in the treatment diarrhea-predominant irritable bowel syndrome.* J Physiol Pharmacol, 2019. **70**: p. 15-24.
- 32. Elmenhorst, D., et al., *Sleep deprivation increases A1 adenosine receptor density in the rat brain.* Brain research, 2009. **1258**: p. 53-58.

- 33. Shimamura, T., et al., *Structure of the human histamine H 1 receptor complex with doxepin.* Nature, 2011. **475**(7354): p. 65-70.
- 34. Zobayer, M., In silico Characterization and Homology Modeling of Histamine Receptors. 2019, Khulna University of Engineering & Technology (KUET), Khulna, Bangladesh.
- 35. Hervig, M.E., et al., Glutamatergic and serotonergic modulation of rat medial and lateral orbitofrontal cortex in visual serial reversal learning. Psychology & Neuroscience, 2020.
- 36. Bortolozzi, A., et al., *The activation of 5-HT2A receptors in prefrontal cortex enhances dopaminergic activity.* Journal of neurochemistry, 2005. **95**(6): p. 1597-1607.
- 37. Furukawa, H., et al., Subunit arrangement and function in NMDA receptors. Nature, 2005. **438**(7065): p. 185-192.
- 38. Bäck, M., et al., *Update on leukotriene, lipoxin and oxoeicosanoid receptors: IUPHAR Review 7.* British journal of pharmacology, 2014. **171**(15): p. 3551-3574.
- 39. Ricciotti, E. and G.A. FitzGerald, *Prostaglandins and inflammation*. Arteriosclerosis, thrombosis, and vascular biology, 2011. **31**(5): p. 986-1000.
- 40. Woodward, D., R. Jones, and S. Narumiya, *International Union of Basic and Clinical Pharmacology. LXXXIII: classification of prostanoid receptors, updating 15 years of progress.* Pharmacological reviews, 2011. **63**(3): p. 471-538.
- 41. McLean, P.G., A. Ahluwalia, and M. Perretti, Association between kinin B1 receptor expression and leukocyte trafficking across mouse mesenteric postcapillary venules. Journal of Experimental Medicine, 2000. **192**(3): p. 367-380.
- 42. Freeman, R.S., et al., NGF deprivation-induced gene expression: after ten years, where do we stand? Progress in brain research, 2004. **146**: p. 111-126.
- 43. Huang, S.M., et al., *An endogenous capsaicin-like substance with high potency at recombinant and native vanilloid VR1 receptors.* Proceedings of the National Academy of Sciences, 2002. **99**(12): p. 8400-8405.
- 44. Sluka, K.A., O.C. Winter, and J.A. Wemmie, *Acid-sensing ion channels: A new target for pain and CNS diseases*. Current opinion in drug discovery & development, 2009. **12**(5): p. 693.

- 45. Förstermann, U. and W.C. Sessa, *Nitric oxide synthases: regulation and function*. European heart journal, 2012. **33**(7): p. 829-837.
- 46. Lorenzo, L.E., et al., Differential organization of γ-aminobutyric acid type A and glycine receptors in the somatic and dendritic compartments of rat abducens motoneurons. Journal of Comparative Neurology, 2007. **504**(2): p. 112-126.
- 47. Iyengar, S., M.H. Ossipov, and K.W. Johnson, *The role of calcitonin gene*related peptide in peripheral and central pain mechanisms including migraine. Pain, 2017. **158**(4): p. 543.
- 48. Edvinsson, L., Role of CGRP in migraine, in Calcitonin Gene-Related Peptide (CGRP) Mechanisms. 2019, Springer. p. 121-130.
- 49. de Bartolomeis, A., et al., *Glycine signaling in the framework of dopamine-glutamate interaction and postsynaptic density. Implications for treatment-resistant schizophrenia.* Frontiers in Psychiatry, 2020. **11**.
- 50. Hashemi, M., S. Bashi, and A. Zali, *The expression level of cannabinoid receptors type 1 and 2 in the different types of astrocytomas.* Molecular biology reports, 2020. **47**(7): p. 5461-5467.
- 51. Abood, M., et al., Cannabinoid receptors (version 2019.4) in the IUPHAR/BPS Guide to Pharmacology Database. IUPHAR/BPS Guide to Pharmacology CITE, 2019. **2019**(4).
- 52. Podlewska, S. and R. Kurczab, Mutual Support of Ligand-and Structure-Based Approaches—To What Extent We Can Optimize the Power of Predictive Model? Case Study of Opioid Receptors. Molecules, 2021. **26**(6): p. 1607.
- Fantinati, A., et al., A diastereoselective synthesis of Cebranopadol, a novel analgesic showing NOP/mu mixed agonism. Scientific reports, 2017. **7**(1): p. 1-7.
- 54. Gan, T., et al. Analgesic and morphine-sparing effects of the peripherally-restricted kappa opioid agonist CR845 after intravenous administration in women undergoing a laparoscopic hysterectomy. in ANESTHESIA AND ANALGESIA. 2013. LIPPINCOTT WILLIAMS & WILKINS 530 WALNUT ST, PHILADELPHIA, PA 19106-3621 USA.
- 55. Vanderah, T.W., et al., *Dynorphin promotes abnormal pain and spinal opioid antinociceptive tolerance*. Journal of Neuroscience, 2000. **20**(18): p. 7074-7079.
- 56. Costantino, C.M., et al., *Opioid receptor heteromers in analgesia*. Expert reviews in molecular medicine, 2012. **14**: p. e9.

- 57. Gerardi, M.C., et al., *Novel pharmaceutical options for treating fibromyalgia*. Expert review of clinical pharmacology, 2016. **9**(4): p. 559-565.
- 58. Choy, E., et al. A systematic review and mixed treatment comparison of the efficacy of pharmacological treatments for fibromyalgia. in Seminars in arthritis and rheumatism. 2011. Elsevier.
- 59. Chan, A.K.M., C.W. Cheung, and Y.K. Chong, *Alpha-2 agonists in acute pain management*. Expert opinion on pharmacotherapy, 2010. **11**(17): p. 2849-2868.
- 60. Medhurst, S.J., et al., *Novel histamine H3 receptor antagonists GSK189254* and GSK334429 are efficacious in surgically-induced and virally-induced rat models of neuropathic pain. Pain, 2008. **138**(1): p. 61-69.
- 61. Ibrahim, M.M., et al., *Activation of CB2 cannabinoid receptors by AM1241 inhibits experimental neuropathic pain: pain inhibition by receptors not present in the CNS.* Proceedings of the National Academy of Sciences, 2003. **100**(18): p. 10529-10533.
- 62. Perret, D. and Z.D. Luo, *Targeting voltage-gated calcium channels for neuropathic pain management*. Neurotherapeutics, 2009. **6**(4): p. 679-692.
- 63. de Lera Ruiz, M. and R.L. Kraus, *Voltage-gated sodium channels:* structure, function, pharmacology, and clinical indications. Journal of medicinal chemistry, 2015. **58**(18): p. 7093-7118.
- 64. Kwong, K. and M.J. Carr, *Voltage-gated sodium channels*. Current opinion in pharmacology, 2015. **22**: p. 131-139.
- 65. Petrenko, A.B., et al., *The role of N-methyl-D-aspartate (NMDA) receptors in pain: a review.* Anesthesia & Analgesia, 2003. **97**(4): p. 1108-1116.
- Parsons, C.G., *NMDA receptors as targets for drug action in neuropathic pain*. European journal of pharmacology, 2001. **429**(1-3): p. 71-78.
- 67. Gavva, N.R., et al., *Pharmacological blockade of the vanilloid receptor TRPV1 elicits marked hyperthermia in humans.* Pain, 2008. **136**(1-2): p. 202-210.
- 68. Andrade, E., F. Meotti, and J. Calixto, *TRPA1 antagonists as potential analgesic drugs*. Pharmacology & therapeutics, 2012. **133**(2): p. 189-204.
- 69. Guan, Y., et al., *Mas-related G-protein-coupled receptors inhibit* pathological pain in mice. Proceedings of the National Academy of Sciences, 2010. **107**(36): p. 15933-15938.

- 70. Bannwarth, B. and M. Kostine, *Targeting nerve growth factor (NGF) for pain management: what does the future hold for NGF antagonists?* Drugs, 2014. **74**(6): p. 619-626.
- 71. Korhonen, T., et al., *The treatment of disc herniation-induced sciatica with infliximab: one-year follow-up results of FIRST II, a randomized controlled trial.* Spine, 2006. **31**(24): p. 2759-2766.
- 72. Landry, R.P., et al., Propentofylline, a CNS glial modulator does not decrease pain in post-herpetic neuralgia patients: in vitro evidence for differential responses in human and rodent microglia and macrophages. Experimental neurology, 2012. **234**(2): p. 340-350.
- 73. Huang, L.J., et al., *Effect of sildenafil on neuropathic pain and hemodynamics in rats.* Yonsei medical journal, 2010. **51**(1): p. 82.
- 74. Von Moos, R., et al., *Metastatic bone pain: treatment options with an emphasis on bisphosphonates*. Supportive Care in Cancer, 2008. **16**(10): p. 1105-1115.
- 75. Klein, R.C., et al., *The Amino Acid Residue at Sequence Position 5 in the Conantokin Peptides Partially Governs Subunit-selective Antagonism of RecombinantN-Methyl-d-aspartate Receptors.* Journal of Biological Chemistry, 2001. **276**(29): p. 26860-26867.
- 76. Adams, D.J., et al., ω-Conotoxin CVID inhibits a pharmacologically distinct voltage-sensitive calcium channel associated with transmitter release from preganglionic nerve terminals. Journal of biological chemistry, 2003. **278**(6): p. 4057-4062.
- 77. Bogin, O., *Venom peptides and their mimetics as potential drugs*. Modulator, 2005. **19**(9): p. 14-20.
- 78. Craik, D., *Venoms to Drugs 2002 Conference*. IDrugs: the investigational drugs journal, 2002. **5**(9): p. 881-884.
- 79. Nilsson, K.P.R., et al., Solution structure of  $\chi$ -conopeptide MrIA, a modulator of the human norepinephrine transporter. Peptide Science: Original Research on Biomolecules, 2005. **80**(6): p. 815-823.

# How to cite this article:

Bhattacharjee B, Deka B and Shkya A. Understanding neurobiological pathways of pain regulation and novel therapeutics for pain management, *Curr Trends Pharm Res*, 2021; 8 (1): 88-118.